

Unveiling the Defensive Role of Flavonoids: A Toxicant Countermeasure

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SUMMARY

Flavonoids are phytochemical substances found in many plants, fruits, vegetables, and leaves, and have potential uses in medicinal chemistry. They are plant-derived secondary metabolites with notable antioxidant capabilities. They are categorized based on their chemical composition, level of unsaturation, and carbon ring oxidation. Flavonoids are classified into many subgroups including anthoxanthins (flavanone and flavanol), flavanones, flavanonols, flavans, chalcones, anthocyanidins, and iso-flavonoids. Each of these flavonoids is ubiquitously found in nature. Consuming more foods rich in flavonoids offers several health advantages. Due to their beneficial impact on human health, there is a growing endeavor to extract these natural substances from different plants. Since these natural compounds have positive effects on human health, an increasing effort has been made to isolate these compounds from various plants. Flavonoids have several medical advantages such as anticancer, antioxidant, anti-inflammatory, and antiviral activities. They possess neuroprotective and cardio-protective properties. The biological effects are influenced by the specific flavonoid type, its potential mechanism of action, and its bioavailability. These affordable pharmaceutical ingredients exhibit notable biological activity and have been shown beneficial in treating many ailments. Recent research has focused on isolating them, creating analogs, and studying their impact on human health using diverse methods and animal models. An increasing number of flavonoids have been identified, with their discovery continuing to grow steadily. Thus, this chapter has endeavored to describe the protective function of flavonoids and their beneficial actions to enhance our comprehension of their impact on human health.

INTRODUCTION

Polyphenols are a large class of biologically active compounds that are present as secondary metabolites in different parts of plants. These compounds provide flavor and color to various parts of plants as well as protect them from environmental toxins owing to their potential biological abilities (Quideau et al., 2011; Daglia, 2012). Flavonoids are common polyphenolic compounds that exhibit a diverse range of biological as well as pharmacological potentials such as anti-cancer, anti-viral, antioxidant, anti-inflammatory, immune modulatory and anti-bacterial properties (Harborne & Williams, 2000). Various investigations revealed the palliative action of flavonoids to counteract various sorts of environmental toxicants. For many decades, pesticide contamination has become a global health concern. The residues of pesticides remain in air, soil, water and sediments through which humans are inevitably exposed to these pollutants (Chrisman et al., 2009). The epidemiological data elucidated that pesticide intoxication exerts deleterious effects on different body organs via endocrine disruption, epigenetic variations, oxidative stress, mitochondrial impairments and autophagy (Mostafalou & Abdollahi, 2013).

Flavonoids are well known for their broad-spectrum ameliorative abilities therefore considered as indispensable components of the pharmaceutical, nutraceutical as well as cosmetic industries. These compounds protect the tissues against oxidative stress owing to their reactive oxygen species (ROS) scavenging potentials (Metodiewa et al., 1997). Free radicals are always present in our bodies in response to normal physiological activity or exogenous damage. It is documented that free radicals directly attack the lipids of plasma membrane which disrupt its normal architecture and trigger a cascade or reaction called lipid peroxidation. The change in permeability alters the osmotic pressure which leads to bursting of cells (Walker et al., 2000). Therefore, flavonoids are considered as major break-through to mitigate the damaging effects of oxidative stress and its associated mechanisms. Various sources of flavonoids and their potential curative roles have been depicted in Fig 1.

The following flavonoids demonstrated marvelous anti-toxicant properties owing to their different attributes.

QUERCETIN

Quercetin is a naturally occurring flavonoid that is widely present in pod vegetables, leaves and fruits of various plants. It is considered as indispensable part of our diet owing to its anti-oxidative, hepato-protective, cardio-protective, reno-protective as well as anti-atherosclerosis potentials (Liu et al., 2010; Cai et al., 2013; Lin et al., 2014). Various organs such as kidneys, liver, small and large intestine are responsible for the metabolism of quercetin. Quercetin is now extensively used as a food supplement as well as a remedial agent against cirrhosis, steatosis and hepatic inflammation. It is revealed that quercetin demonstrates its antioxidant abilities owing to its chemical structural configuration which enable it to donate free electrons and scavenge free radicals (Ramya & Padma, 2013).

Numerous investigations reported that quercetin potentially counteracts mycotoxins induced toxicities which are attributed to its antioxidant and anti-inflammatory abilities. Quercetin escalates the activities of antioxidant enzymes, reduces oxidative stress as well as prevents the onset of lipid peroxidation (Saleem et al., 2015). Quercetin ameliorates aflatoxin prompted oxidative stress and cytotoxicity via regulating Nrf2/keap1 pathways. Furthermore, quercetin supplementation prevents DNA damage as well as micronucleus owing to its antigenic abilities. Similarly, Quercetin administration inhibits ochratoxin instigated cytotoxicity and significantly reduces the levels of pro-inflammatory cytokines (Schoneberg et al., 2018). Moreover, quercetin reduces oxidative stress and prevents the onset of ischemia and reperfusion in cardiomyocytes via regulating xanthine oxidase system (Sanhuesa et al., 1992).

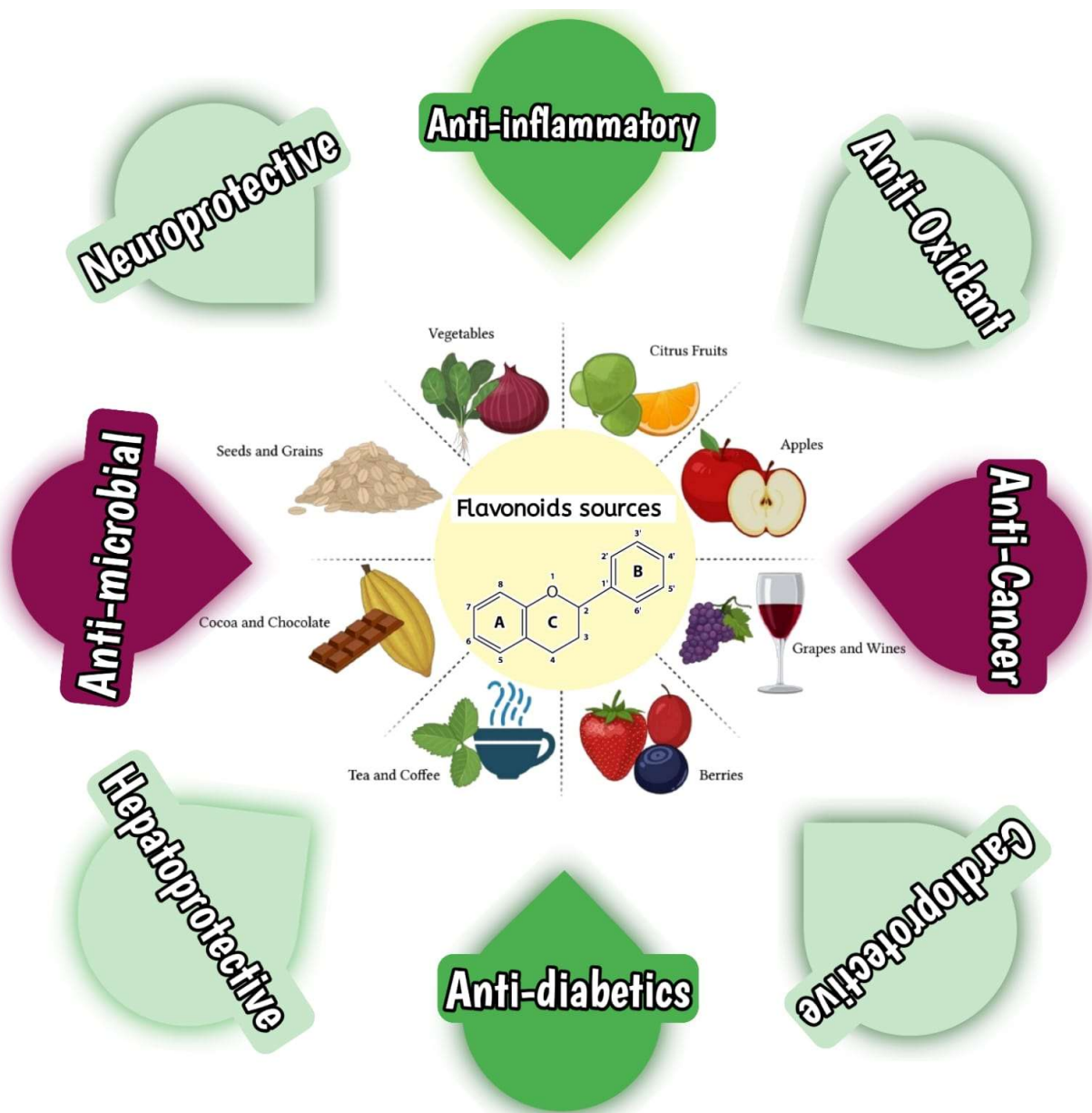


Fig 1. Some sources of flavonoids and their potential role

Aforementioned findings indicate that Quercetin is a potential therapeutic compound to avert various toxicities.

KAEMPFEROL

Kaempferol is an essential flavonoid that is found in various plants such as beans, kale, leek, grapes and broccoli. Numerous investigations have reported anti-microbial, antioxidative, anti-obesity, neuroprotective, cardioprotective and anti-cancerous properties. Rats treated with kaempferol showed a prominent reduction in the levels of inflammatory cytokines (Yeh et al., 2022). Recent investigation has documented the hepatoprotective abilities of kaempferol. It is revealed that kaempferol treatment remarkably reduced the liver function markers in CCl₄ induced hepatotoxicity in rats (Zang et al., 2017). Furthermore, kaempferol prevents the alcohol prompted liver damage via reducing the expression of CYP2E1 while escalating the expression of cytoprotective antioxidant genes (Wang et al., 2015). Moreover, kaempferol binds to ALK5 which triggers the downregulation of TGF- β /Smads pathway thereby preventing the liver against fibrosis (Xu et al., 2019). Lipopolysaccharides (LPS) are potential compounds that trigger the development of hepatic fibrosis. LPS activates TLR4 receptors in hepatic and Kupffer cells (Singh et al., 2017). Furthermore, it triggers the activation of pro-inflammatory cytokines in hepatic tissues (Cheng et al., 2017). Additionally, LPS induces nitrosative as well as oxidative stress which ultimately reduces antioxidant activities and damages the liver (Latha et al., 2017). However, kaempferol administration reduces the expression of TLR4 receptor which ultimately inhibits the activation of pro-inflammatory cytokines particularly NF- κ B in hepatic tissues (Islam et al., 2019). These findings elucidate that kaempferol can be used as a therapeutic agent against various toxicities. However, clinical trials should be conducted to evaluate the efficacy of kaempferol on human beings.

RUTIN

Rutin is classified as a flavonoid glycoside and is a part of the broader class of plant compounds known as flavonoids. This compound is found in various fruits, vegetables, and plant-based diets, contributing not only to their potential health benefits. Rutin is abundant in citrus fruits, buckwheat, asparagus, onions, and tea. Its consumption has been associated with several health-promoting effects, making it a subject of interest in both traditional medicine and modern scientific research (Chua, 2013). Rutin has been used widely in the field of medicine due to its numerous pharmacological properties including anti-allergic, vasoactive, anti-inflammatory, anti-bacterial, anti-tumor, anti-viral, and anti-protozoal properties (Calabrò et al., 2005). In addition, hypolipidemic and cytoprotective activities have also been reported (Casa et al., 2000). Due to its potent antioxidant properties and high radical scavenging activity, rutin has antiplatelet, antiviral and anti-hypertensive qualities along with strength in blood vessel capillaries. The stability of the genome and the prevention of disease are two possible benefits of these characteristics. Based on certain research, rutin antioxidant properties in the Fenton reaction (Caillet et al., 2007) and its ability to inhibit

low-density lipoprotein (LDL) peroxidation are dose-dependent (Jiang et al., 2007).

LUTEOLIN

Luteolin is a natural flavonoid that is found in many edible plants such as broccoli, celery, chrysanthemum flower, parsley and onion leaves, and (Lim et al., 2013). Luteolin acts as both a pro-oxidant and an antioxidant biochemically and it demonstrates a variety of biological benefits including anti-inflammatory, anti-allergy, and anti-cancer properties. In traditional Chinese medicine, plants that are high in luteolin have been used to treat conditions like cancer inflammatory disorders, and hypertension. The biological effects of luteolin may be connected functionally; for instance, the anti-inflammatory action may be connected to the cancer-preventing properties of luteolin. (Lin et al., 2008). In addition, anti-diabetic, cytoprotective (Kempuraj et al., 2021) and anti-angiogenic (Raffa et al. 2017), properties have also been reported. Furthermore, luteolin exhibits the ability to penetrate the blood-brain barrier and function as a neuroprotective agent (Theoharides et al., 2016). Researchers reported that LUT inhibits neuroinflammatory and systemic responses in coronavirus disease (Kempuraj et al., 2021). Luteolin demonstrates potent cardiovascular protection via intricate pathways that transmit signals and targeting effectors, which aligns with recent developments in our understanding of oxidative stress and inflammatory mechanisms of the cardiovascular system. A high luteolin diet probably lowers the risk of sudden cardiac arrest (Si et al., 2014). According to recent research, mast cell-induced neuroinflammation was inhibited by the phytosomal formulation of LUT, which reduced long-term COVID syndrome related to brain fog (Theoharides et al., 2021). In isolated peripheral blood mononuclear cells from human MS (multiple sclerosis) patients, luteolin may also have therapeutic immunomodulatory effects when used in separation. Moreover, pro-inflammatory cytokines like interleukin-1 β (IL-1 β), tumor necrosis factor-alpha (TNF- α), and the ratio of the cell migration mediator MMP-9 to its inhibitor TIMP-1 showed additive effects when luteolin treatment was administered (Sternberg et al., 2009).

MYRICETIN

Myricetin is a flavonoid that is abundant in natural plants belonging to various families, such as the Fagaceae, Vitaceae, Rosaceae, Leguminosae, Primulaceae, Myricaceae, Ericaceae and Compositae. It is frequently found as an essential active component and additive in many foods including vegetables, fruits, berries, merlot, tea and honey (Nardini & Garaguso, 2020). Further, it demonstrated a wide range of pharmacological properties, including antidiabetic, analgesic, antitumor, hepatoprotective, and anti-inflammatory effects. When related to other flavonoids, myricetin's three hydroxyl groups on ring B are thought to contribute to its strong antioxidant action. Research conducted in vitro has demonstrated that the two hydroxyl groups and the double-bonded oxygen group are responsible for the mineral chelation effect (Wu et al., 2013).

Myricetin's prooxidative impact is caused by the catechol groups in its structure, which result in the formation of semi-quinone radicals. When the 4-hydroxyl group on the C ring and the 4-hydroxyl group on the B ring combine to produce a quinone, this radical gets oxidized. Myricetin has been linked to numerous health advantages, including lowered hepatic triglyceride levels, prevention of liver damage, less oxidative stress, and lower cholesterol levels. The hepatoprotective properties of myricetin have been extensively studied by numerous experts worldwide (Semwal et al., 2016). Myricetin possesses various pharmacological properties against organ damage as shown in Fig 2.

CHRYSIN

Chrysin (CH) is a vital flavonoid that is prevalently present in numerous plant compositions, propolis and honey (Eldutar et al. 2017). According to recent investigations, CH possesses the potential to exhibit various medicinal and commercial properties such as anti-inflammatory (Kandemir et al. 2017a), antioxidant (Eldutar et al. 2017), anti-apoptotic, anti-diabetic, anti-cancerous, anxiolytic, anti-allergic (Du et al. 2012) and anti-estrogenic activities. Chrysin also acts as an anti-aging agent against D-galactose induced aging (Anand et al., 2012). Chrysin plays significant protective role against agent induced colitis(Dextran sodium sulfate, Dimethylhydrazine, cisplatin), neurotoxicity (Lipopolysaccharide, Streptozotocin, formalin) nephrotoxicity (Tetrachlorodibenzo-p-dioxin, Doxorubicin, Fluorouracil, ethanol, cisplatin, adenine, Carbon tetrachloride), respiratory toxicity(Ovalbumin, Bleomycin, Cigarette smoke), hepatotoxicity (Tert-butyl hydroperoxide, ethanol, Methotrexate), cardiovascular toxicity [Triton WR-1339, Streptozotocin, Doxorubicin, Mitoxantrone (Pingili et al., 2019)].

MTX is an efficient chemotherapeutic agent, its clinical use is limited due to its toxic effects on the liver (Hagag et al., 2015). Chrysin was discovered to have a protective role against the hepatotoxic effects of MTX by lowering the markers of liver injury (Ali et al., 2014). Cyclophosphamide (CYP) is widely used in Chemotherapy and also an alkylating drug of the oxazaphosphorin family (Ahlmann & Hempel, 2016). Due to its side effects including vomiting, alopecia, bone marrow suppression, nephrotoxicity, neurotoxicity, nausea, urotoxicity, hepatotoxicity, immunotoxicity and cardiotoxicity the clinical usage of this medication is restricted (Taslimi et al. 2019). According to some reports, chrysin is a very potent flavonoid that has a wide range of pharmacological actions, including inhibiting the activity of topoisomerases that deoxyribonucleic acid (Russo et al., 2012), anti-inflammatory activity by preventing the release of histamine and pro-inflammatory cytokine expression (Bae et al., 2011), and anticancer activity by supporting the degradation of caspase-8 and caspase 3 by TRAIL (Samarghandian et al., 2011).

The inhibitory impact on vascular endothelial growth factor (VEGF) instigated angiogenesis (Tsuji & Walle, 2007; Li et al., 2011), cardioprotective activity by enhancing post-ischemic functional recovery (Tian et al., 2014), inhibition of (HDACs) (Szkudelski, 2001), anti-asthmatic activity by suppressing NF-κB and stimulate nitric oxide synthase

(Wadibhasme et al., 2011), inhibition of TNF-α and interleukin (IL)-1β (Bai et al., 2013), prevention of programmed cell death by initiation of estrogen receptor (Zeng et al., 2013), Reno-protective activity by glucose-induced renal tubular cell migration with diminishing matrix metalloproteinase-2 activity (Kang et al., 2015), antihypertensive (Farkhondeh et al., 2015), antidiabetogenic (Samarghandian et al., 2016), anti-hypercholesterolemic action (Anandhi et al., 2014), and stopping breast cancer cells from spreading metastatically (Lirdprapamongkol et al., 2013).

TANGERETIN

Bisphenol A, Benzo[a]pyrene (BaP) and Potassium dichromate (PD) are different toxicants that induce toxicity. TNG is a natural bioactive Polymethoxy flavones (in which all hydroxyl groups are capped by methyl residues) extracted from peel tissues of citrus species including citrus depressa and citrus tangerine. TNG processes neuroprotective, antioxidant, anti-inflammatory, anti-cancerous and anti-diabetogenic actions. Additionally, TNG play a substantial role in various pharmacological activities. The anti-cancer properties of TNG are attributed to their metabolic resistance, high membrane permeability and variety of biological activities, including cell growth inhibitory actions. It is anticipated that they will serve as biotherapeutic medications to preserve human health by aiding in the inhibition and cure of severe ailments like neurodegenerative diseases, lipid metabolism disorders, cancer and neurological diseases (Nahar et al., 2012).

BPA is an industrialized toxin that may harm the liver. The United States Environmental Protection Agency (EPA) has designated the BPA a third most harmful environmental toxicant TNG's antioxidant and anti-inflammatory potential significantly prevent liver damage caused by BPA. TGN's pharmacological abilities enable it to decrease ROS generation and increase antioxidant enzyme activity BPA can slow down organismal aging. Dietary TAN supplementation restored the transcript levels of heat shock proteins and mitigated the BaP-induced loss in motility, pumping, and poly-Q accretion (Roth et al., 2014).

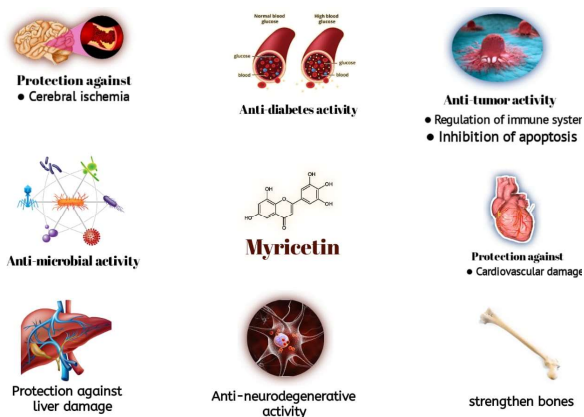


Fig 2. Demonstration of various pharmacological properties of Myricetin against organ damages

Environmental xenobiotic potassium dichromate (PD) is well known to be carcinogenic, mutagenic and teratogenic to both animals and humans. According to reports, TNG act as neuro-protective drug which protects the brain from damage caused by PD. TNG also reduced the expression of caspase-3 in the brain. TNG also plays a major neuroprotective role by regulating the (Nrf2) nuclear factor erythroid 2-related factor 2 signaling pathway, preventing the production of inflammatory mediators (interleukins, tumor necrosis factor and interferons), and inhibiting programmed cell death in response to PD-induced acute brain injury (Sharma et al., 2020).

Nrf2 has a cytoprotective and neuroprotective response through the regulation of several antioxidant genes. Recent research has shown that Nrf2 is upregulated in brain injury as an endogenous defensive response. Apoptosis is a program of heritably regulated cell death which leads to internucleosomal DNA fragmentation and has a major influence on the pathophysiology of brain injury. In many illnesses of the central nervous system (CNS), initiation of caspase-3 is an eminent feature of neurodegeneration, and this pathway is thought to be a prospective target for therapy (Macks et al. 2022). Moreover, oxidative stress is a major component that triggers caspase-3-mediated apoptotic pathways. Numerous studies claim that by lowering neuroinflammation in experimental animals, TNG mitigates cognitive and memory impairments (Yu et al., 2020). Research showed that the brain could be shielded from chromium-induced brain damage by activating the antioxidant pathways and inhibiting pro-apoptotic and inflammatory mediators (Xu et al. 2022).

MORIN

Morin, a natural flavonoid derived from the Moraceae family has antioxidant, anti-inflammatory, anti-carcinogenic, neuroprotective, antidiabetic and antiproliferative properties. It has been found to alter the Nrf2 pathway in research. It has a systemic protective effect, lowering the harmful aftereffects of various medications deprived of interrupting with their functioning. It is thus regarded as a strong beneficial medication for all illnesses caused mostly by free radical damage (Kuzu M. et al. 2018).

Another research looked at whether morin protects against testicular toxicity induced by ifosfamide (IFOS), a drug used to treat a diversity of malignancies. The findings acquired demonstrated that IFOS accomplished oxidative stress in testicular tissues (Ozturk et al. 2014). Oxidative stress has been reported to induce ERS, irritation, apoptosis, DNA damage and autophagy. Morin therapy reduces oxidative stress. It reduced TNF- α and IL-1 β protein levels, indicating anti-inflammatory activity (Miltonprabu et al., 2017). It also elevated mRNA transcript levels for the ERS markers ATF-6, PERK, IRE1, GRP-78, and CHOP, as well as the apoptotic markers Bax, Casp-3, and apaf-1. It increased expression of the anti-apoptotic protein Bcl-2 and the cell survival signal AKT-2 genes. Morin demonstrated an anti-autophagic action and decreased the levels of the protein beclin-1. Morin also reduced the quantity of 8-OHdG immune-positive cells and mitigated oxidative DNA damage (Cakmak et al., 2023).

The toxicity of cyclophosphamide (CPX) severely restricts its usage as an immunosuppressive or anti-cancer medication. Significant alterations are seen, such as in the hematological system, particularly in the numbers of leukocytes and platelets (Azevedo et al. 2007). Citrus extract, for instance, has been demonstrated to lessen CPX-induced genotoxicity in mouse bone marrow cells, especially in relation to its flavonoid contents with antioxidative activity (Hosseinimehr & Karami, 2005). The above-mentioned study found that adding morin to the treatment decreased the amount of body weight lost.

PELARGONIDIN

The pelargonidin is important flavonoid based on anthocyanins and is utilized in medications to treat skeletal myopathy, retinopathy, hypoglycemia, and other conditions. Studies demonstrate the effectiveness of anthocyanins in avoiding a range of chronic illnesses, including obesity, diabetes, and cardiovascular disease (Li et al., 2015). Based on their anti-inflammatory, antioxidant, anti-mutagenic, and antiangiogenic properties, besides their ability to induce cell differentiation, inhibit proliferation through signal transduction pathway modification, tempt cell cycle arrest, and promote autophagy in cancerous cells, anthocyanins may have antitumor effects. Strawberries and food items with red coloring are the primary sources of pelargonidin (Praveena, R. et al. 2022).

Glycosyl flavonoids, including pelargonidin, chromenylum-3, glycosides, oxane-3,4,5-triol, are absorbed by the body and metabolized throughout the digestive process. Acetylated forms of pelargonidin glycosides were discovered in urine samples from experimental animals and people, according to studies on anthocyanins' absorption and metabolism (Karlsen, A. et al. 2007). Both pelargonidin (2-(4-hydroxyphenyl) chromenylum-3,5,7-triol)—(C₁₅H₁₁O₅⁺) and pelargonidin 3-O- β -glucopyranoside, also referred to as pelargonidin-3-Oglucoside—(C₂₁H₂₁O₁₀⁺)—have been utilized as food additive substitutes as a result of their antioxidant characteristics. Due to their proven ability to cause hyperglycemia, chromosomal abnormality, and DNA damage, these chemicals are recognized to constitute a major risk to human health (Habtemariam, 1997; Karlsen, A. et al. 2010).

Measurements of permeability, stimulation of pro-inflammatory proteins, adherence of neutrophil and migration in LPS-activated human umbilical vein endothelial cells (HUVECs) and mice were used to assess the anti-inflammatory properties of PEL (Mantawyet al., 2014). It was discovered that PEL prevented the rupture of the barrier caused by LPS, the production of CAMs, and the adherence and trans-endothelial relocation of neutrophils to endothelial cells of human (Li et al., 2011). Additionally, LPS-tempted hyper permeability and leukocyte exodus were inhibited in vivo by the impact of PEL on the barrier reliability of HUVECs was assessed using a permeability test (Miltonprabu et al., 2017). The barrier integrity remained unchanged after treatment with PEL (30 μ M). The HUVECs were subjected to different PEL concentrations for six hours following the injection of LPS (100 ng/mL) for four hours. The outcomes indicate PEL the findings show that PEL prevents endothelial cells from experiencing LPS-mediated hyperpermeability; the best

effects are obtained at concentrations greater than 10 μM (Berman R.S., et al. 1993; (Ginis Z et. al. 2016).

It has been noted that the rings of pelargonidin are covered with electron accepting π^* type ring road, sometimes referred to as lowest unoccupied molecular orbitals (LUMO) (Lee et al., 2017). The highest occupied molecular orbital (HOMO), or electron-donating π type orbitals, are discovered to be distributed throughout the entire molecular expanse, mostly above the B-ring, in case of flavonoids serves as the primary reactive or donor position Goldblum et al., 1993; Alvarez-Suarez et al., 2014). This finding unequivocally demonstrates that the carbonyl carbon and hydroxyl entities of the B-ring in pelargonidin play an electron-donating role which facilitate it to play its ROS scavenging properties (Ashrafizadeh et al., 2020).

CONCLUSION

Flavonoids are a diverse group of molecules that occur naturally in several plants, including fruits and vegetables, as well as in plant-derived products like coffee, chocolate, and tea. Flavonoids have been consistently documented to provide a variety of health advantages. These phytochemicals are abundant in antioxidants, offering our body innate immunological defenses against everyday external and internal poisons. It is strongly suggested to include various kinds of flavonoids into our daily diet to promote health and minimize the risk of serious illnesses including diabetes mellitus, cancer, stroke, and heart attack. Flavonoids have shown therapeutic benefits in most preclinical trials conducted on mouse models. Various groups of flavonoids have been identified with important biological properties including anticancer, antibacterial, antifungal, anti-diabetic, antimarial, neuroprotective, cardio-protective, and anti-inflammatory effects. Thus, it is deduced that above-mentioned flavonoids compounds could be used as therapeutic candidates to cure several ailments.

REFERENCES

- Ahlmann M & G Hempel, 2016. The effect of cyclophosphamide on the immune system: Implications for clinical cancer therapy. *Cancer Chemotherapy and Pharmacology* 78:661-71. <https://doi.org/10.1007/s00280-016-3152-1>
- Ali N, S Rashid, S Nafees et al., 2014. Beneficial effects of chrysin against methotrexate-induced hepatotoxicity via attenuation of oxidative stress and apoptosis. *Molecular and Cellular Biochemistry* 285:215-23. <https://doi.org/10.1007/s11010-013-1830-4>
- Alvarez-Suarez JM, F Giampieri, S Tulipani et al., 2014. One-month strawberry-rich anthocyanin supplementation ameliorates cardiovascular risk, oxidative stress markers and platelet activation in humans. *The Journal of Nutritional Biochemistry* 25:289-94. <https://doi.org/10.1016/j.jnutbio.2013.11.002>
- Anandhi R, MSM Jaabir, PA Thomas et al., 2012. Protective role of chrysin against oxidative stress in d-galactose-induced aging in an experimental rat model. *Geriatrics and Gerontology International* 12:741-50. <https://doi.org/10.1111/j.1447-0594.2012.00843.x>
- Anandhi R, PA Thomas & P Geraldine, 2014. Evaluation of the anti-atherogenic potential of chrysin in Wistar rats. *Molecular and Cellular Biochemistry* 385:103-13. <https://doi.org/10.1007/s11010-013-1819-z>
- Ashrafizadeh M, Z Ahmadi, R Mohammadinejad et al., 2020. Tangeretin: A mechanistic review of its pharmacological and therapeutic effects. *Journal of Basic and Clinical Physiology and Pharmacology* 31:20190191. <https://doi.org/10.1515/jbcpp-2019-0191>
- Azevedo L, PL de Lima, JC Gomes et al., 2007. Differential response related to genotoxicity between eggplant (*Solanum melanogena*) skin aqueous extract and its main purified anthocyanin (delphinidin) in vivo. *Food and Chemical Toxicology* 45:852-8. <https://doi.org/10.1016/j.fct.2006.11.004>
- Bae Y, S Lee & SH Kim, 2011. Chrysin suppresses mast cell-mediated allergic inflammation: Involvement of calcium, caspase-1 and nuclear factor- κB . *Toxicology and Applied Pharmacology* 254:56-64. <https://doi.org/10.1016/j.taap.2011.04.008>
- Bai J, Y Luo, Z Song et al., 2013. Effects and the mechanisms of chrysin on sepsis-associated acute lung injury of rats chrysin inhibits acute lung injury. *Life Science Journal* 10:1052-8.
- Berman RS, JD Frew & W Martin, 1993. Endotoxin-induced arterial endothelial barrier dysfunction assessed by an in vitro model. *British Journal of Pharmacology* 110:1282-4. <https://doi.org/10.1111/j.1476-5381.1993.tb13956.x>
- Cai X, Z Fang, J Dou et al., 2013. Bioavailability of quercetin: Problems and promises. *Current Medicinal Chemistry* 20:2572-82. <https://doi.org/10.2174/09298673113209990120>
- Caillet S, H Yu, S Lessard et al., 2007. Fenton reaction applied for screening natural antioxidants. *Food Chemistry* 100:542-52. <https://doi.org/10.1016/j.foodchem.2005.10.009>
- Cakmak F, S Kucukler, C Gur et al., 2023. Morin provides therapeutic effect by attenuating oxidative stress, inflammation, endoplasmic reticulum stress, autophagy, apoptosis, and oxidative DNA damage in testicular toxicity caused by ifosfamide in rats. *Iranian Journal of Basic Medical Sciences* 26:1227-36.
- Calabro ML, S Tommasini, P Donato et al., 2005. The rutin/ β -cyclodextrin interactions in fully aqueous solution: Apectroscopic studies and biological assays. *Journal of pharmaceutical and biomedical analysis* 36:1019-27. <https://doi.org/10.1016/j.jpba.2004.09.018>
- Casa LC, I Villegas, DL Lastra et al., 2000. Evidence for protective and antioxidant properties of rutin, a natural flavone, against ethanol induced gastric lesions. *Journal of Ethnopharmacology* 71:45-53. [https://doi.org/10.1016/S0378-8741\(99\)00174-9](https://doi.org/10.1016/S0378-8741(99)00174-9)
- Cheng P, T Wang, W Li et al., 2017. Baicalin alleviates lipopolysaccharide-induced liver inflammation in chicken by suppressing TLR4-mediated NF- κB pathway. *Frontiers in Pharmacology* 8:547-612. <https://doi.org/10.3389/fphar.2017.00547>
- Chrisman JDR, S Koifman, PdN Sarcinelli et al., 2009. Pesticide sales and adult male cancer mortality in Brazil. *International Journal of Hygiene and Environmental Health* 212:310-21. <https://doi.org/10.1016/j.ijheh.2008.07.006>
- Chua LS, 2013. A review on plant-based rutin extraction methods and its pharmacological activities. *Journal of Ethnopharmacology* 150:805-17. <https://doi.org/10.1016/j.jep.2013.10.036>
- Daglia M, 2012. Polyphenols as antimicrobial agents. *Current Opinion in Biotechnology* 23:174-81. <https://doi.org/10.1016/j.copbio.2011.08.007>
- Du Q, X Gu, J Cai et al., 2012. Chrysin attenuates allergic airway inflammation by modulating the transcription factors T-bet and GATA-3 in mice. *Molecular Medicine Reports* 6:100-4.
- Eldutar E, FM Kandemir, S Kucukler et al., 2017. Restorative effects of Chrysin pretreatment on oxidant-antioxidant status, inflammatory cytokine production, and apoptotic and autophagic markers in acute paracetamol-induced hepatotoxicity in rats: an experimental and biochemical study. *Journal of Biochemical and Molecular Toxicology* 31:21960. <https://doi.org/10.1002/jbt.21960>
- Farkhondeh T, S Samarghandia, M Azimin-Nezha et al., 2015. Effect of chrysin on nociception in formalin test and serum levels of noradrenalin and corticosterone in rats. *International Journal of Clinical and Experimental Medicine* 8:2465-70.
- Ginis Z, G Ozturk, A Albayrak et al., 2016. Protective effects of caffeic acid phenethyl ester on ifosfamide-induced central neurotoxicity in rats. *Toxicology and Industrial Health* 32:337-43. <https://doi.org/10.1177/0748233713500817>
- Goldblum SE, X Ding, TW Brann et al., 1993. Bacterial lipopolysaccharide induces actin reorganization, intercellular gap formation, and endothelial barrier dysfunction in pulmonary vascular endothelial cells: Concurrent F-actin depolymerization and new actin synthesis. *Journal of Cellular Physiology* 157:13-23. <https://doi.org/10.1002/jcp.1041570103>
- Habtemariam S, 1997. Flavonoids as inhibitors or enhancers of the cytotoxicity of tumor necrosis factor- α in L-929 tumor cells. *Journal of Natural Products* 60:775-8. <https://doi.org/10.1021/np960581z>
- Hagag AA, AM AbdElal, MS Elfarag et al., 2015. Therapeutic value of black seed oil in methotrexate hepatotoxicity in Egyptian children with acute lymphoblastic leukemia. *Infectious Disorders-Drug Targets* 15:64-71. <https://doi.org/10.2174/1871526515666150320161440>
- Harborne JB & CA Williams, 2000. Advances in flavonoid research since 1992. *Phytochemistry* 55:481-504. [https://doi.org/10.1016/S0031-9422\(00\)00235-1](https://doi.org/10.1016/S0031-9422(00)00235-1)

- Hosseinimehr SJ & M Karami, 2005. Citrus extract modulates genotoxicity induced by cyclophosphamide in mice bone marrow cells. *Journal of Pharmacy and Pharmacology* 57:505-9. <https://doi.org/10.1211/0022357055849>
- Islam MS, H Yu, L Miao et al., 2019. Hepatoprotective effect of the ethanol extract of *Illicium henryi* against acute liver injury in mice induced by lipopolysaccharide. *Antioxidants* 8:446-521. <https://doi.org/10.3390/antiox8100446>
- Jiang P, F Burczynski, C Campbell et al., 2007. Rutin and flavonoid contents in three buckwheat species *Fagopyrum esculentum*, *F. tataricum*, and *F. homotropicum* and their protective effects against lipid peroxidation. *Food Research International* 40:356-64. <https://doi.org/10.1016/j.foodres.2006.10.009>
- Kandemir F, S Kucukler, E Eldutar et al., 2017. Chrysin protects rat kidney from paracetamol-induced oxidative stress, inflammation, apoptosis, and autophagy: A multi-biomarker approach. *Scientia Pharmaceutica* 85:4. <https://doi.org/10.3390/scipharm85010004>
- Kang MK, SH Park, YJ Choi et al., 2015. Chrysin inhibits diabetic renal tubulointerstitial fibrosis through blocking epithelial to mesenchymal transition. *Journal of Molecular Medicine* 93:759-72. <https://doi.org/10.1007/s00109-015-1301-3>
- Karlsen A, I Paur, SK Bohn et al., 2010. Bilberry juice modulates plasma concentration of NF- κ B related inflammatory markers in subjects at increased risk of CVD. *European Journal of Nutrition* 49:345-55. <https://doi.org/10.1007/s00394-010-0092-0>
- Karlsen A, L Retterstøl, P Laake et al., 2007. Anthocyanins inhibit nuclear factor- κ B activation in monocytes and reduce plasma concentrations of pro-inflammatory mediators in healthy adults. *The Journal of Nutrition* 137:1951-4. <https://doi.org/10.1093/jn/137.8.1951>
- Kempuraj D, R Thangavel, DD Kempuraj et al., 2021. Neuroprotective effects of flavone luteolin in neuroinflammation and neurotrauma. *Biofactors* 47:190-7. <https://doi.org/10.1002/biof.1687>
- Kuzu M, FM Kandemir, S Yildirim et al., 2018. Morin attenuates doxorubicin-induced heart and brain damage by reducing oxidative stress, inflammation and apoptosis. *Biomedicine and Pharmacotherapy* 106:443-53. <https://doi.org/10.1016/j.biopha.2018.06.161>
- Latha S, S Chaudhary & R Rs, 2017. Hydroalcoholic extract of *Stevia rebaudiana* bert. leaves and stevioside ameliorates lipopolysaccharide induced acute liver injury in rats. *Biomedicine and Pharmacotherapy* 95:1040-50. <https://doi.org/10.1016/j.biopha.2017.08.082>
- Lee MH, HJ Cha, EO Choi et al., 2017. Antioxidant and cytoprotective effects of morin against hydrogen peroxide-induced oxidative stress are associated with the induction of Nrf-2 mediated HO-1 expression in V79-4 Chinese hamster lung fibroblasts. *International Journal of Molecular Medicine* 39:672-80. <https://doi.org/10.3892/ijmm.2017.2871>
- Li D, Y Zhang, Y Liu et al., 2015. Purified anthocyanin supplementation reduces dyslipidemia, enhances antioxidant capacity, and prevents insulin resistance in diabetic patients. *The Journal of Nutrition* 145:742-8. <https://doi.org/10.3945/jn.114.205674>
- Li X, JN Wang, JM Huang et al., 2011. Chrysin promotes tumor necrosis factor (TNF)-related apoptosis-inducing ligand (TRAIL) induced apoptosis in human cancer cell lines. *Toxicology in Vitro* 25:630-5. <https://doi.org/10.1016/j.tiv.2010.12.013>
- Lim SH, Sk Jung, S Byun et al., 2013. Luteolin suppresses UVB-induced photoaging by targeting JNK1 and p90RSK2. *Journal of Cellular and Molecular Medicine* 17:672-80. <https://doi.org/10.1111/jcmm.12050>
- Lin SY, YY Wang, WY Chen et al., 2014. Beneficial effect of quercetin on cholestatic liver injury. *The Journal of Nutritional Biochemistry* 25:1183-95. <https://doi.org/10.1016/j.jnutbio.2014.06.003>
- Lin Y, R Shi, X Wang et al., 2008. Luteolin, a flavonoid with potential for cancer prevention and therapy. *Current Cancer Drug Targets* 8:634-46. <https://doi.org/10.2174/156800908786241050>
- Lirdprapamongkol K, H Sakurai, S Abdelhamed et al., 2013. A flavonoid chrysin suppresses hypoxic survival and metastatic growth of mouse breast cancer cells. *Oncology Reports* 30:2357-64. <https://doi.org/10.3892/or.2013.2667>
- Liu CM, YL Zheng, J Lu et al., 2010. Quercetin protects rat liver against lead-induced oxidative stress and apoptosis. *Environmental Toxicology and Pharmacology* 29:158-66. <https://doi.org/10.1016/j.etap.2009.12.006>
- Macks C, D Jeong, S Bae et al., 2022. Dexamethasone-loaded hydrogels improve motor and cognitive functions in a rat mild traumatic brain injury model. *International Journal of Molecular Sciences* 23:11153. <https://doi.org/10.3390/ijms231911153>
- Mantawy EM, WM El-Bakly, A Esmat et al., 2014. Chrysin alleviates acute doxorubicin cardiotoxicity in rats via suppression of oxidative stress, inflammation and apoptosis. *European Journal of Pharmacology* 728:107-18. <https://doi.org/10.1016/j.ejphar.2014.01.065>
- Metodiewa D, A Kochman & S Karolczak, 1997. Evidence for antiradical and antioxidant properties of four biologically active N, N-Diethylaminoethyl ethers of flavone oximes: A comparison with natural polyphenolic flavonoid rutin action. *IUBMB Life* 41:1067-75. <https://doi.org/10.1080/15216549700202141>
- Miltonprabu S, M Tomczyk, K Skalicka-Wozniak et al., 2017. Hepatoprotective effect of quercetin: From chemistry to medicine. *Food and Chemical Toxicology* 108:365-74. <https://doi.org/10.1016/j.fct.2016.08.034>
- Mostafalou S & M Abdollahi, 2013. Pesticides and human chronic diseases: Evidence, mechanisms, and perspectives. *Toxicology and Applied Pharmacology* 268:157-77. <https://doi.org/10.1016/j.taap.2013.01.025>
- Nahar MS, AS Soliman, JA Colacino et al., 2012. Urinary bisphenol A concentrations in girls from rural and urban Egypt: A pilot study. *Environmental Health* 11: 1-8. <https://doi.org/10.1186/1476-069X-11-20>
- Nardini M & I Garaguso, 2020. Characterization of bioactive compounds and antioxidant activity of fruit beers. *Food Chemistry* 305:125437. <https://doi.org/10.1016/j.foodchem.2019.125437>
- Ozturk G, Z Gimis, SN Kurt et al., 2014. Effect of alpha lipoic acid on ifosfamide-induced central neurotoxicity in rats. *International Journal of Neuroscience* 124:110-6. <https://doi.org/10.3109/00207454.2013.823962>
- Pingili RB, AK Pawar, SR Challa et al., 2019. A comprehensive review on hepatoprotective and nephroprotective activities of chrysin against various drugs and toxic agents. *Chemico-Biological Interactions* 308:51-60. <https://doi.org/10.1016/j.cbi.2019.05.010>
- Praveena R, A Balasankar, K Aruchamy et al., 2022. Structural Activity and HAD inhibition efficiency of pelargonidin and its glucoside-a theoretical approach. *Molecules* 27:8016. <https://doi.org/10.3390/molecules27228016>
- Quideau S, D Deffieux, C Douat-Casassus et al., 2011. Plant polyphenols: Chemical properties, biological activities, and synthesis. *Angewandte Chemie International Edition* 50:586-621. <https://doi.org/10.1002/anie.201000044>
- Raffa D, B Maggio, MV Raimondi et al., 2017. Recent discoveries of anticancer flavonoids. *European Journal of Medicinal Chemistry* 142:213-28. <https://doi.org/10.1016/j.ejmech.2017.07.034>
- Ramya P & VV Padma, 2013. Ochratoxin-induced toxicity, oxidative stress and apoptosis ameliorated by quercetin-Modulation by Nrf2. *Food and Chemical Toxicology* 62:205-16. <https://doi.org/10.1016/j.fct.2013.08.048>
- Roth S, MR Spalinger, I Müller et al., 2014. Bilberry-derived anthocyanins prevent IFN- γ -induced pro-inflammatory signalling and cytokine secretion in human THP-1 monocytic cells. *Digestion* 90:179-89. <https://doi.org/10.1159/000366055>
- Russo P, AD Bufalo & A Cesario, 2012. Flavonoids acting on DNA topoisomerases: recent advances and future perspectives in cancer therapy. *Current Medicinal Chemistry* 19:5287-93. <https://doi.org/10.2174/092986712803833272>
- Salem IB, A Prola, M Boussabeh et al., 2015. Crocin and Quercetin protect HCT116 and HEK293 cells from Zearalenone-induced apoptosis by reducing endoplasmic reticulum stress. *Cell Stress and Chaperones* 20:927-38. <https://doi.org/10.1007/s12192-015-0613-0>
- Samarghandian S, JT Afshari & S Davoodi, 2011. Chrysin reduces proliferation and induces apoptosis in the human prostate cancer cell line pc 3. *Clinics* 66:1073-9. <https://doi.org/10.1590/S1807-59322011000600026>
- Samarghandian S, M Azimi-Nezhad, F Samini et al., 2016. Chrysin treatment improves diabetes and its complications in liver, brain, and pancreas in streptozotocin-induced diabetic rats. *Canadian Journal of Physiology and Pharmacology* 94:388-93. <https://doi.org/10.1139/cjpp-2014-0412>
- Sanhueza J, J Valdes, R Campos et al., 1992. Changes in the xanthine dehydrogenase/xanthine oxidase ratio in the rat kidney subjected to ischemia-reperfusion stress: Peffect of some flavonoids. *Research Communications in Chemical Pathology and Pharmacology* 78:211-8.
- Schöneberg T, K Kibler, M Sulyok et al., 2018. Can plant phenolic compounds reduce Fusarium growth and mycotoxin production in cereals? *Food Additives and Contaminants* 35:2455-70. <https://doi.org/10.1080/19440049.2018.1538570>
- Semwal DK, RB Semwal, S Combrinck et al., 2016. Myricetin: A dietary molecule with diverse biological activities. *Nutrients* 8:90. <https://doi.org/10.3390/nu8020090>
- Sharma V, A Kaur & TG Singh, 2020. Counteracting role of nuclear factor erythroid 2-related factor 2 pathway in Alzheimer's disease. *Biomedicine and Pharmacotherapy* 129:110373. <https://doi.org/10.1016/j.biopha.2020.110373>
- Si H, RP Wyeth, & D Liu, 2014. The flavonoid luteolin induces nitric oxide production and arterial relaxation. *European Journal of Nutrition* 53:269-75. <https://doi.org/10.1007/s00394-013-0525-7>
- Singh A, C Koduru, Carlisle et al., 2017. NADPH oxidase 4 modulates hepatic responses to lipopolysaccharide mediated by Toll-like receptor-4. *Scientific Reports* 7:1-12. <https://doi.org/10.1038/s41598-017-14574-8>
- Sternberg Z, K Chadha, A Lieberman et al., 2009. Immunomodulatory responses of peripheral blood mononuclear cells from multiple sclerosis patients upon in vitro incubation with the flavonoid luteolin: Additive effects of IFN- β . *Journal of Neuroinflammation* 6:1-8. <https://doi.org/10.1186/1742-2094-6-28>

- Szkudelski T, 2001. The mechanism of alloxan and streptozotocin action in B cells of the rat pancreas. *Physiological Research* 50:537-46.
- Taslimi P, FM Kandemir, Y Demir et al., 2019. The antidiabetic and anticholinergic effects of chrysin on cyclophosphamide-induced multiple organ toxicity in rats: Pharmacological evaluation of some metabolic enzyme activities. *Journal of Biochemical and Molecular Toxicology* 33:22313. <https://doi.org/10.1002/jbt.22313>
- Theoharides TC, C Cholevas, K Polyzoidis et al., 2021. Long-COVID syndrome-associated brain fog and chemofog: Luteolin to the rescue. *Biofactors* 47:232-41. <https://doi.org/10.1002/biof.1726>
- Theoharides TC, I Tsilioni, AB Patel et al., 2016. Atopic diseases and inflammation of the brain in the pathogenesis of autism spectrum disorders. *Translational Psychiatry* 6:844. <https://doi.org/10.1038/tp.2016.77>
- Tian SS, FS Jiang, K Zhang et al., 2014. Flavonoids from the leaves of *Carya cathayensis* Sarg. inhibit vascular endothelial growth factor-induced angiogenesis. *Fitoterapia* 92:34-40. <https://doi.org/10.1016/j.fitote.2013.09.016>
- Tsuji PA & T Walle, 2007. Benzo [a] pyrene-induced cytochrome P450 1A and DNA binding in cultured trout hepatocytes-inhibition by plant polyphenols. *Chemico-Biological Interactions* 169:25-31. <https://doi.org/10.1016/j.cbi.2007.05.001>
- Wadibhasme PG, MM Ghaisas & PA Thakurdesai, 2011. Anti-asthmatic potential of chrysin on ovalbumin-induced bronchoalveolar hyperresponsiveness in rats. *Pharmaceutical Biology* 49:508-15. <https://doi.org/10.3109/13880209.2010.521754>
- Walker EH, ME Pacold, O Perisic et al., 2000. Structural determinants of phosphoinositide 3-kinase inhibition by wortmannin, LY294002, quercetin, myricetin, and staurosporine. *Molecular Cell* 6:909-19. [https://doi.org/10.1016/S1097-2765\(00\)00088-5](https://doi.org/10.1016/S1097-2765(00)00088-5)
- Wang M, J Sun, Z Jiang et al., 2015. Hepatoprotective effect of kaempferol against alcoholic liver injury in mice. *The American Journal of Chinese Medicine* 43:241-54. <https://doi.org/10.1142/S0192415X15500160>
- Wu t, Z Yu, Q Tang et al., 2013. Honeysuckle anthocyanin supplementation prevents diet-induced obesity in C57BL/6 mice. *Food and Function* 4:1654-61. <https://doi.org/10.1039/c3fo60251f>
- Xu M, W Wang, W Lu et al., 2022. Evodiamine prevents traumatic brain injury through inhibiting oxidative stress via PGK1/NRF2 pathway. *Biomedicine and Pharmacotherapy* 153:113435. <https://doi.org/10.1016/j.biopha.2022.113435>
- Xu T, S Huang, Q Huang et al., 2019. Kaempferol attenuates liver fibrosis by inhibiting activin receptor-like kinase 5. *Journal of Cellular and Molecular Medicine* 23:6403-10. <https://doi.org/10.1111/jcmm.14528>
- Yeh TM, CD Chang, SS Liu et al., 2022. Tea seed kaempferol triglycoside attenuates LPS-induced systemic inflammation and ameliorates cognitive impairments in a mouse model. *Molecules* 27:2055. <https://doi.org/10.3390/molecules27072055>
- Yu S, X Liu, D Yu et al., 2020. Morin protects LPS-induced mastitis via inhibiting NLRP3 inflammasome and NF-κB signaling pathways. *Inflammation* 43:1293-303. <https://doi.org/10.1007/s10753-020-01208-x>
- Zang Y, D Zhang, C Yu et al., 2017. Antioxidant and hepatoprotective activity of kaempferol 3-O-β-D-(2, 6-di-O-α-L-rhamnopyranosyl) galactopyranoside against carbon tetrachloride-induced liver injury in mice. *Food Science and Biotechnology* 26:1071-6. <https://doi.org/10.1007/s10068-017-0170-7>
- Zeng W, Y Yan, F Zhang et al., 2013. Chrysin promotes osteogenic differentiation via ERK/MAPK activation. *Protein and Cell* 4:539-47. <https://doi.org/10.1007/s13238-013-3003-3>